Proton, Carbon-13, and Nitrogen-15 Nuclear Magnetic Resonance Studies of [¹⁵N]Azoles: 1-Phenylpyrazole and the Tautomerically Mobile 3-Methyl-1-phenylpyrazolin-5-one

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The ¹H, ¹³C, and ¹⁵N chemical shifts and ¹H–¹³C, ¹H–¹⁵N, ¹³C–¹⁵N, and ¹⁵N–¹⁵N coupling constants of 95% ¹⁵Nenriched 1-phenylpyrazole and 1-phenyl-3-methylpyrazolin-5-one have been determined and assigned. In the case of 3-methyl-1-phenyl[¹⁵N₂]pyrazolin-5-one only the ¹⁵N n.m.r. spectrum [solvent (CD₃)₂SO] shows that slow exchange occurs between the NH and OH tautomers.

A RECENT survey of the literature concerning the tautomerism of heterocycles ¹ shows that ¹H n.m.r. spectroscopy has proved a very useful technique. However ¹³C n.m.r. spectroscopy is being more and more utilised to compete with and even to replace proton resonance. Although it is recognised ^{2,3} that ¹⁵N n.m.r. spectroscopy may furnish structural information, the method is seldom used because ¹⁵N-labelled compounds are necessary if ¹⁵N-¹⁵N or ¹³C-¹⁵N coupling constants are to be measured.

The availability of phenyl[$^{15}N_2$]hydrazine led us to synthesise two new labelled compounds: 1-phenyl[$^{15}N_2$]-pyrazole, as a reference molecule, and 3-methyl-1-phenyl[$^{15}N_2$]pyrazolin-5-one, the object of our interest; the tautomerism of this molecule has been studied previously ¹ and it is thus appropriate for a study of the applicability of ^{15}N n.m.r. to tautomeric problems.

1-Phenyl[¹⁵N₂]pyrazole (1).—The ¹³C chemical shifts and ¹³C-¹⁵N couplings for 1-phenyl[¹⁵N₂]pyrazole are summarised in Table 1. Assignments of the ¹³C resonances are the same as quoted by Stothers ⁴ from the data of Rees and Green,⁵ however the assignments of the couplings to N-1 or N-2 are not so certain. The observed coupling of C-3 (1.2 Hz) could be to either N-1 or N-2 since each of these couplings is expected to be small: ¹ $J_{\rm CN}$ in pyridine ⁶ is 0.45 Hz, ¹ $J_{\rm CN}$ in quinoline ⁷ is 0.6—2.4 Hz, and ² $J_{\rm CN}$ in pyrrole ⁸ is 4 Hz. Similarly

⁸ S. Bulusu, J. R. Autera, and T. Axenrod, in 'Nuclear Magnetic Resonance Spectroscopy of Nuclei Other Than Protons,' eds. T. Axenrod and G. A. Webb, Wiley, New York and London, 1974, ch. 7.

1974, ch. 7. ⁴ J. B. Stothers, 'Carbon-13 NMR Spectroscopy,' Academic Press, New York and London, 1972, ch. 7. it is not possible to assign the two couplings at C-4 specifically to N-1 or N-2. The large coupling (12.1 Hz) at C-5 must be to N-1 by comparison with the values 12.8 Hz for ${}^{1}J_{\rm CN}$ in pyrrole,⁸ 12.0 Hz for the pyridinium ion,⁶ and 13.8 and 15.9 Hz for the quinolinium ion ⁷ (${}^{1}J_{\rm CN}$ for a trigonal nitrogen atom has the smaller value noted above for pyridine and quinoline only when the



nitrogen atom has a lone pair). For C-2' the two couplings are equal, and the doublet splitting (2.0 Hz) of C-3' is probably the three-bond coupling to N-1 (cf. ${}^{3}J_{CN}$ in aniline 9 is 1.2 Hz).

The ¹⁵N chemical shifts, ¹ $J_{\rm NN}$ values, and ¹H–¹⁵N coupling constants for compound (1) are also given in Table 1. The N-1 signal is assigned as that to high field (by 82 p.p.m.) by analogy with the reported ¹⁰ ¹⁴N assignments for *N*-methylpyrazole. The value for the one-bond ¹⁵N–¹⁵N coupling (12.8 Hz) is within the range noted by Bulusu *et al.*³ (4.5–19 Hz). The ¹H–¹⁵N coupling constants obtained from the proton spectrum of com-

⁵ R. G. Rees and M. J. Green, J. Chem. Soc. (B), 1968, 387.
 ⁶ R. L. Lichter and J. D. Roberts, J. Amer. Chem. Soc., 1971,

⁶ R. L. Lichter and J. D. Roberts, J. Amer. Chem. Soc., 1971, 93, 5218.
 ⁷ P. S. Pregosin, E. W. Randall, and A. I. White, J.C.S.

⁷ P. S. Pregosin, E. W. Randall, and A. I. White, *J.C.S. Perkin II*, 1972, 1.

⁸ J. M. Briggs, E. Rahkamaa, and E. W. Randall, J. Magnetic Resonance, 1973, 11, 416.
⁹ A. I. White, Ph.D. Thesis, University of London, 1972.

A. I. White, Ph.D. Thesis, University of London, 1972.
 M. Witanowski, L. Stefaniak, H. Januszewski, Z. Grabowski,

and G. A. Webb, Tetrahedron, 1972, 28, 637.

¹ J. Elguero, C. Marzin, A. R. Katritzky, and P. Linda, 'The Tautomerism of Heterocycles,' Academic Press, New York and London, 1976.

and London, 1976. ² M. Witanowski, L. Stefaniak, and H. Januszewski, in 'Nitrogen NMR,' eds. M. Witanowski and G. A. Webb, Plenum Press, London and New York, 1973, ch. 4. ³ S. Bulusu, J. R. Autera, and T. Axenrod, in 'Nuclear

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pound (1) are similar in magnitude to those for $[^{15}N]$ pyrrole,¹¹ 5-phenyl[¹⁵N]isothiazole,¹² [¹⁵N]pyrazole,¹³ and ¹⁵N isoxazole.¹⁴ In all these previous studies it was concluded that the ¹H-¹⁵N coupling constants were negative, as must be the case here for compound (1).

3-Methyl-1-phenyl[¹⁵N₂]pyrazolin-5-one (2).—This tautomeric compound has been studied previously in dimethyl sulphoxide solution by Feeney et al.¹⁵ by ¹³C n.m.r. (¹⁵N at natural abundance). Our ¹³C data for the di-15N-labelled material in CDCl₃ are given in Table 2. In this solvent the compound exists predominantly as tautomer a, as evidenced by the highfield resonance for C-4 and by a triplet pattern in the ¹H-(continuous wave) decoupled ¹³C spectrum, characteristic of the -CH₂- group. The ¹³C resonances of the phenyl ring were assigned by using the 1-phenylpyrazole assignments (Table 1) and off-resonance ¹H (continuous wave) decoupling experiments.

As with 1-phenylpyrazole, the assignment of the

TABLE 1

¹³C and ¹⁵N N.m.r. parameters for 1-phenyl[¹⁵N₂]pyrazole ^a

lucleus	8 "	Multiplicity	$J(^{13}C-^{15}N)/Hz$
C-3	141.1	d	1.2 + 0.3
C-4	107.6	d, d	2.1 ± 0.3 ,
			6.2 ± 0.3
C-5	126.8	d	12.1 ± 0.3 (N-1)
C-1'	٥ 140.5		
C-2'	119.4	t	$1.6~\pm~0.3$
C-3′	129.5	d	$2.0~\pm~0.3$ (N-1)
C-4′	126.5	s	,
N-1	198.4	d	12.8 ± 1.2 ^d
N-2	280.4	d	$12.8~\pm~1.2$ d

⁶ 0.9M-Solution in CDCl₃. ^b ¹³C Chemical shifts in p.p.m. downfield from internal Me₄Si; ¹⁵N chemical shifts in p.p.m. downfield from ammonium ion reference (see Experimental section). "This resonance was not observed from the 15Nenriched sample, presumably because of the multiplicity and a long spin-lattice relaxation time reducing the signal to noise ratio. The chemical shift was measured from a sample containing ¹⁵N at the natural abundance level. ^d This value is ${}^{1}J({}^{15}N-$ ¹⁵N). In addition the following parameters were measured The 100 MHz proton spectrum: δ (H-3) 7.69, δ (H-4) 6.40, δ (H-5) 7.85; ${}^{2}J_{H(5)-N(1)}$ 4.4, ${}^{3}J_{H(3)-N(1)}$ 7.4, ${}^{3}J_{H(4)-N(1)}$ 6.0, ${}^{2}J_{H(3)-N(2)}$ 14.2, ${}^{3}J_{H(4)-N_{2}}$ 1.0 Hz.

observed ¹³C-¹⁵N couplings to N-1 or N-2 is complicated. The assignments of the ¹³C-¹⁵N couplings at the phenyl ring carbon atoms follow those for compound (1). Lichter et al.¹⁶ have studied ¹³C-¹⁵N couplings in some aliphatic amides in which they found ${}^{1}J_{CN}$ (to the carbonyl carbon atom) in the range 13.4-15.1 Hz and $^{2}J_{\rm CN}$ (across the carbonyl group) in the range 6.9–10.3 Hz. Accordingly we assign the larger splittings at C-5 (11.0 Hz) and C-4 (13.3 Hz) to N-1. Thus the two-bond is larger than the one-bond coupling. As with 1phenylpyrazole, the lack of available correlations precludes definite assignment of the 3.1 Hz splitting at C-3 to either N-1 or N-2. The large (9.8 Hz) splitting at the methyl carbon atom (C-6) is due to N-2. As the

¹¹ E. Rahkamaa, Z. Naturforsch., 1969, 24a, 2004.

¹² D. Crepaux and J. M. Lehn, Mol. Phys., 1968, **14**, 547; J. P. Kintzinger and J. M. Lehn, Chem. Comm., 1967, 660.

¹³ J. P. Jacobsen, O. Snerling, E. J. Pedersen, J. T. Nielsen, and K. Shaumburg, J. Magnetic Resonance, 1973, 10, 130.
 ¹⁴ D. Crepaux and J. M. Lehn, Org. Magnetic Resonance, 1975,

7, 524.

basis for this assignment we use the data of Lichter et al.,¹⁷ who found ${}^{2}J_{CN}$ for anti-oxime types to be 2.4—11.6 Hz and for syn-oxime types to be 1-2 Hz.

The ¹⁵N chemical shifts and ${}^{1}J_{NN}$ values for compound (2) are given in Table 2. As for 1-phenylpyrazole

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¹³C and ¹⁵N N.m.r. parameters for 3-methyl-1-phenyl-¹⁵N₂]pyrazolin-5-one (2) in CDCl₂^a

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Nucleus	8 0	Multiplicity	$J(^{18}C-^{15}N)/Hz$
C-3	156.2	d	3.1 + 0.6
C-4	43.0	d, d	13.3 ± 0.6 (N-1),
			1.5 ± 0.6 (N-2)
C-5	170.6	d, d	11.0 ± 0.6 (N-1),
			< 1.2 (N-2)
C-6	16.8	d	9.8 ± 2.4 (N-2)
C-1′	138.3	d	12.2 ± 1.2 (N-1)
C-2'	118.9	t	$1.2~\pm~0.3$
C-3′	128.8	d	1.7 ± 0.3 (N-1)
C-4′	125.0	s	
N-1	172.2	d	12.0 ± 0.3 °
N-2	304.2	d	12.0 ± 0.3 °

^a 0.7M-Solution. ^b See footnote b Table 1. ^c This value is ${}^{1}J({}^{15}N-{}^{15}N)$. In addition ${}^{3}J_{H(6)-N(2)}$ (3.5 Hz) was measured from the 100 MHz proton spectrum.

(Table 1), the N-1 signal is assigned as that at higher field.

Proton n.m.r. studies ¹ of the pyrazolone (unlabelled) in Me₂SO have indicated a major change in the position of the tautomeric equilibrium in comparison with solutions in chloroform. In Me₂SO solution 20% of



compound (2) exists in the form a, slowly interconverting with tautomers b and c. Tautomers b and c are in fast equilibrium; usually 1 it is believed that tautomer b predominates. Our ¹³C n.m.r. data for the pyrazolone (2) in Me₂SO are summarised in Table 3. The assignments follow those of Feeney et al.¹⁵ except that we have interchanged the C-3' and -5' assignment with that of C-4' by considering the relative intensities of the resonances. It is noteworthy that the resonance from C-4 in $(CD_3)_2$ SO solution is substantially to lower field, as expected from b or c, than when observed from a solution in CDCl₃ (Table 2). The assignment of this peak to a -CH- group was confirmed by its doublet nature in the

¹⁵ J. Feeney, G. A. Newman, and P. J. S. Pauwels, *J. Chem. Soc.* (C), 1970, 1842. ¹⁶ R. L. Lichter, C. G. Fehder, P. H. Patton, J. Combes, and

D. E. Dorman, J.C.S. Chem. Comm., 1974, 114.

¹⁷ R. L. Lichter, D. E. Dorman, and R. Wasylishen, J. Amer. Chem. Soc., 1974, 96, 930.

off-resonance ¹H-(continuous wave) decoupled spectrum. Two small resonances observed at 118.1 and 124.5 p.p.m. were assigned to C-2' and C-4' of the minor tautomer a, in slow exchange with the major form $b \Longrightarrow c$. The characteristic resonance from C-4 of a was obscured by the solvent resonance. The resolution obtained in the ¹³C spectrum of compound (2) in $(CD_a)_2$ SO was poorer

TABLE 3 ¹³C N.m.r. parameters for 3-methyl-1-phenyl¹⁵N₂]pyrazolin-5-one (2) in $(CD_3)_2 \overline{SO}^a$ 80 Carbon Multiplicity /(13-15N)/Hz C-3 148.4 $\begin{array}{c} 9.8 \ \pm \ 2.4 \ (\text{N-1}) \\ 15.9 \ \pm \ 1.2 \ (\text{N-1}) \\ 7.3 \ \pm \ 2.4 \ (\text{N-2}) \\ 18.3 \ \pm \ 2.4 \ (\text{N-1}) \end{array} \right)$ C-4 C-5 C-6 C-1' C-2' C-3' 89.0 d 154.6d d 13.9 Major form d 138.4120.4s 128.8s 124.9 s C-2 C-4 118.1 s Minor form 124.5s ^a 0.7M-Solution. ^b See footnote b Table 1

than for the solution in CDCl_3 , with the result that only the larger ${}^{13}\text{C}_{-16}\text{N}$ couplings (given in Table 3) were measurable. These couplings were assigned as for the solution in CDCl_3 .

The ¹⁵N n.m.r. spectrum of compound (2) in $(CD_3)_2SO$ has proved crucial for the discussion (see Figure). It



¹H Noise-decoupled ¹⁵N spectra of 3-methyl-1-phenyl[¹⁵N₂] pyrazolin-5-one (2) in $(CD_3)_2SO$; (i) at 30 °C; 14 000 pulses; (ii) at 85 °C; 8 192 pulses; X refers to a probable decomposition product. These are both 'magnitude' spectra and so contain no information on the sign of the ¹⁵N-{¹H} nuclear Overhauser enhancement

took a much longer accumulation time than did the spectrum of (2) in CDCl_3 (form *a*). Two well resolved doublets were observed at 299.9 and 167.8 p.p.m. and a weak absorption at 171.6 p.p.m. In addition there is a broad (Δv_4 ca. 125 Hz) resonance at about 234 p.p.m.

* The spectrum in $CDCl_s$ similarly showed the lower field of the two doublets to be substantially the more intense. This may be due to a combination of a differential ¹⁵N-¹H nuclear Overhauser enhancement or different spin-lattice relaxation times for the two nitrogen nuclei. The low-field doublet and the absorption at 171.6 p.p.m. are due to the minor component (a),* whereas the doublet at 167.8 p.p.m. and the broad resonance must be due to the combination of tautomers b and c interconverting at some intermediate rate. To further investigate this proposal, the ¹⁵N spectrum of (2) in $(CD_3)_2SO$ was measured at 85 °C (see Figure). The broad resonance disappeared and two new absorptions were apparent, at 223.5 (doublet) and 158.2 p.p.m. Measurement of the spectrum again at 30 °C showed that the doublet at 223.5 p.p.m. reverted to the broad resonance, whereas the resonance at about 158 p.p.m. persisted. We therefore feel that this latter resonance is due to a decomposition product.

If tautomers b and c are interconverting at some intermediate rate (R) then the sharp nature of the higher field doublet (due to N-1) shows that it is in the fast exchange limit at 30 °C, whereas the broad nature of the N-2 resonance indicates that it is not, and that Ris comparable to the frequency separation between the resonance positions for N-2 in b and c. In order to substantiate this proposition it is necessary to estimate the ¹⁵N chemical shifts for tautomers b and c.

For tautomer b we use as a starting point the ¹⁵N shifts for the pyrazole (1), and introduce corrections for the methyl and hydroxy-substituents. We have taken the nitrogen chemical shift substituent corrections from the data quoted by Witanowski *et al.* on the methyl-substituted pyrazoles ¹⁸ and hydroxy-substituted pyridines.² Thus for N-2 in b we take the value 280 p.p.m. from (1) and correct first by 5 p.p.m. (upfield) for the methyl group two bonds removed and then by 22 p.p.m. (upfield) for the hydroxy-group three bonds removed. This yields the prediction of 253 p.p.m. for N-2 in b. Similarly the shift of N-1 in b is predicted to be 140 p.p.m.

For tautomer c we note that N-1 is in a similar chemical environment to N-1 of a (observed at 171.6 p.p.m.). The difference in the chemical environment of N-2 between b and c may be compared with the difference for the two tautomers of 4-hydroxypyridine (which is believed to exist predominantly as the keto form). The calculated ² nitrogen chemical shift for the hydroxyform is 97 p.p.m. to lower field than the observed shift. If we take this as the difference for N-2 between b and c, then the N-2 resonance of c is predicted to occur at about 156 p.p.m. The observed and calculated ¹⁵N shifts for tautomers b and c are summarised in Table 4.

Although the calculated ¹⁵N shifts are not very precise, the arguments above are confirmed. First, the observed average ¹⁵N shifts do lie between the extremes calculated for b and c. Secondly, the calculated ¹⁵N chemical shift separation between b and c is greater for N-2 (97 p.p.m. = 885 Hz) than for N-1 (32 p.p.m. = 292 Hz).

That the ^{15}N spectrum of compound (2) in $(CD_3)_2SO$ displays broadening due to the exchange between b and c, whereas the ^{13}C spectrum does not, requires

¹⁸ M. Witanowski, L. Stefaniak, H. Januszewski, and J. Elguero, J. Chim. phys., 1973, 70, 697.

comment. The ¹³C resonance likely to show the greatest chemical shift difference between b and c is that due to C-5. Feeney et al.¹⁵ have measured the ¹³C chemical shifts for two model compounds, (3) and (4), closely related to tautomers b and c, respectively. The measured ¹⁵ ¹³C chemical shifts (converted to the Me₄Si scale) are as indicated, and these must be corrected in the following manner. Stothers ¹⁹ concludes that the



¹³C chemical shift substituent effect due to a hydroxygroup upon an sp^2 carbon atom is essentially the same as upon an sp^3 carbon atom. If we assume the same holds true for the ethoxy-substituent [in (3)] then we can predict the ¹³C shift of C-5 in b. Formal conversion of ethanol into diethyl ether shifts the α -13C resonance 20 downfield by 10.1 p.p.m. Thus we predict the chemical shift of C-5 of b to occur at 145.8 p.p.m. Considering compound (4), we note that the carbonyl ¹³C shifts ¹⁹ for 2- and 3-methylcyclopent-2-enone are within 0.4 p.p.m. of the shift for cyclopent-2-enone. Therefore the effect of the N-methyl substituent in (4) is small at C-5, and (4) is a good model for tautomer c: the C-5 signal is predicted to occur at about 166.8 p.p.m.

Thus the predicted ¹³C shift difference for C-5 between b and c is 21 p.p.m. = 475 Hz. This value is substantially less than the 885 Hz shift difference for N-2

TABLE 4					
Estimated values for $K = [b]/[c]$ from ¹³ C and ¹⁵ N					
chemical shifts of compound (2) in $(CD_3)_2SO$					

Chemical shift						
served	Calculated	Calcula				

	Observed	Calculated	Calculated	K =
Nucleus	average b,c	ь	с	[b]/[c]
C-5	154.6	145.8	166.8	1.39
N-1	167.8	140	172	0.15
N-2	223.5	253	156	2.29

and explains why the ¹³C spectrum does not display any resonance broadening due to the chemical exchange.

We believe the ¹³C chemical shifts predicted above are more reliable than the predicted ¹⁵N shifts. The equilibrium constant, K = [b]/[c], calculated from the various predicted and observed chemical shifts is given in Table 4. The values of K derived * from C-5 and N-2 data are in reasonable agreement (1.39 and 2.29)

* In the solid state 3-methyl-1-phenylpyrazolin-5-one has been shown ²¹ to exist as a 50 : 50 mixture of tautomers b and c.

20 Ch. 5 of ref. 4.

whereas that from N-1 (0.15) is notably different. The most likely source for the discrepancy is in the calculation of the shift for N-1 of tautomer b employing the large hydroxy-substituent effect (-57 p.p.m.).

Conclusion .- The observation of temperature-dependent ¹⁵N spectra of the pyrazolone (2) in (CD₃)₂SO leads us to conclude that all three tautomers (a-c) are present. The reason why this is apparent from ¹⁵N spectra rather than from ¹³C spectra is because the ¹⁵N chemical shift difference (in Hz) for the N-2 site (the site of protonation) is much greater than for any of the ¹³C sites (which are all at least one bond removed from the site of protonation). The appearance of the ¹⁵N spectrum at 30 °C leads to the conclusion that the equilibrium between the tautomers is best represented as:

$$a \xrightarrow{\text{Step 1 or 3}} [b \xrightarrow{\text{Step 2}} c]$$

where the activation parameter for step 2 is lower than for step 1 or 3.

Previous proton n.m.r. data ¹ showing 20% of (2) in the form a, coupled with the deduction here from ^{13}C chemical shift data that [b]/[c] is ca. 1.4, give the following overall contributions to the structure of (2) at ca. 30 °C in Me₂SO solution: a, 20%; b, 47%; c, 33%.

EXPERIMENTAL

N.m.r. Spectra.—¹³C (22.63 MHz) and ¹⁵N (9.12 MHz) spectra were obtained with a Bruker HFX-13 instrument operating in the pulse-Fourier transform mode. The instrument was equipped with a B-SV-2 ¹H broad band decoupler unit and free induction decays were accumulated in a Fabritek 1074 CAT (4 K store) and transformed with a PDP-8/I computer. The digitisation in the resulting frequency domain spectra was 2.44 Hz per channel (5 kHz spectral width) or 0.244 Hz per channel (500 Hz spectral width). Pulse flip angles of about 30° were employed for both ¹³C and ¹⁵N spectra, without a 'pulse delay.' ¹³C Spectra were referenced to Me_4Si and ^{15}N spectra to the $^{15}\mathrm{NH_4^{+}}$ resonance from external $^{15}\mathrm{NH_4^{+15}NO_3^{-}}$ (5m in 2N-HNO₃). The samples were contained in 10 mm o.d. tubes and deuterium in the solvent provided the fieldfrequency stabilisation signal.

¹H Spectra were obtained with a Varian HA-100 instrument operating in the frequency sweep mode.

Materials.—Phenyl[¹⁵N₂]hydrazine (95% enrichment) was obtained from Isokommerz. 1-Phenyl¹⁵N₂]pyrazole was prepared by the method of Finar and Hurlock ²² (65% yield) and 3-methyl-1-phenyl[15N2]pyrazolin-5-one was prepared according to Knorr 23 but by heating for 1 h at 100-110 °C instead of 15 min (60% yield).

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²¹ F. Bechtel, J. Gaultier, and C. Hauw, Cryst. Struct. Comm., 1973, 2, 469.
 ²² I. L. Finar and R. J. Hurlock, J. Chem. Soc., 1957, 3024.

23 L. Knorr, Ber., 1883, 16, 2597.

¹⁹ Ch. 8 of ref. 4.